TOTAL SYNTHESIS OF TYLONOLIDE, AN AGLYCONE OF TYLOSIN

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Summary: The total synthesis of tylonolide, a 16-membered-ring aglycone of a macrolide antibiotic, tylosin, has been accomplished by coupling two segments of Cl-Cl0 and Cll-Cl7 portions, which are stereospecifically derived from D-glucose.

Josamycin (leucomycin A₃) and tylosin (1) constitute the medicinally important 16membered-ring macrolide antibiotics. The first total synthesis of josamycin has previously been reported in these laboratories. 1 Herein, as an extension of the strategy, we report the first total synthesis of tylonolide $(\lambda \beta)$, an aglycone of tylosin (λ) .² The construction of the CI-CIO segment of χ_{β} began with the preparation of the acetonide $\frac{\alpha}{2}$ ([α], -17°) from the previously described 3-C-methyl-D-glucoside λ in 42% overall yield in 4 steps: (1) Ac $_2$ O/H $_2$ SO $_4$ 20°; (2) MeONa/MeOH, 20°; (3) Me₂CO/CuSO₄/H₂SO₄, 35°; (4) aq. AcOH, 60°. Periodate oxidation (NaIO₄/90% MeOH, 20°, 1 h) of $\frac{1}{2}$ followed by Wittig reaction ([Ph₃PCH₂OCH₃]Cl⁵/DMS0 anion⁶/THF, 20°, 0.5 h) gave the enol ether, which was hydrolyzed with 90% AcOH to afford the aldehyde 5^3 (92%, $\lbrack \alpha \rbrack_n$ -15°). Reduction (NaBH₄/MeOH, 20°) of ζ followed by successive benzylation (BzlBr/ NaH/DMF, 20° , 1 h) and hydrolysis (5% H₂SO₄/t-BuOH, 50°) afforded the 5-deoxyfuranose β^3 (50%, $[\alpha]_D$ +43°). Reaction of ξ with Ph₃P=CHCOOMe (PhMe, 50°, 18 h) effected Wittig coupling to the E-unsaturated ester, which upon treatment with 2,2-dimethoxypropane (TsOH/DMF, 20°, 1 h) afforded the acetonide \int_0^∞ (73%, $[\alpha]_{\rm p}$ +13°). The introduction of an aldehyde equivalent into the C6 position $^\prime$ was effected by Michael addition as previously described. 1 Reaction of $\vec{\chi}$ with a reagent prepared from 1.5 equiv each of FAMSO 8 and BuLi (-78°, 2.5 h) gave the two adducts $\begin{array}{c} \texttt{8} \\ \texttt{8} \end{array}$ and $\begin{array}{c} \texttt{8} \\ \texttt{8} \end{array}$ (TLC, Rf 0.22 and 0.20, respectively, PhH–Me $_2$ CO 4 : 1), which were separated by column chromatography to give $\beta \lambda^3$ (39%, [α]₀ +25°) and $\beta \lambda^3$ (40%, [α]_n -8°). Since both adducts are converted into the same aldehyde $\lambda \lambda$ as described later on, the production of two adducts is found to be attributed to the asymmetry at C7, not at C6. Their C6 positions in question are considered to have the right configuration required for the synthesis (as depicted by $\{a_i, b_i\}$ from the similar possibility of forming a metal chelate to that previously described¹ and, finally, the successful completion of the synthesis presented below. Reaction of δ with EtSH (BF₃-Et₂0, 201, 18 h) produced the ethyl methyl dithioacetals β and β with lactonization: ϑ ² (35%, [α]_D +51°, Rf 0.28 on TLC with PhH-EtOAc 4 : 1); ϑ ^b (37%, [α]_D +3°, Rf 0.25). The other $\frac{8}{2}$ also gave the aforesaid dithioacetals $\frac{9}{2}$ and $\frac{9}{2}$ on the same procedure, indicating that the formation of the epimers $9a$ and $9b$ was attributable to the newly produced asymmetry at C7. Reduction of \mathfrak{A} and \mathfrak{A} with DIBAL (PhMe, -78°, 0.5 h) afforded the corresponding hemiacetals which were treated with Amberlyst 15 (H type) in MeOH (20°, 2 h) to give the methyl furanosides $\frac{1}{2}$ and $\frac{1}{2}$, respectively: $\frac{1}{2}$ $\frac{3}{2}$ (80%, [α]_n +110°, Rf 0.35 on TLC with PhH-EtOAc 4 : l); \downarrow Q $_\mathrm{R}^\mathrm{D}$ (80%, [α]_n +68°, Rf 0.32), each of which was shown to consist of only one anomer by the NMR. Benzylation (BzlBr/NaH/DMF, 20°, 4 h) of λ Q_R followed by removal of the thioacetal group (CdCO₃/HgCl₂/80% Me₂CO, 20°, 1 h) afforded the labile aldehyde $\lambda\lambda^3$ (85%), which, on standing, was epimerized to the thermodynamically preferred E-isomer 11^{1^3} . Also,

J&Q gave the aforesaid JJ by the same procedure. The Wlttig reaction of ,Q wrth the previously described ylid 12 $\,$ (PhMe, 60 $^{\circ}$, 90 h) occurred stereospecifically to give the Zunsaturated ketone $13^{1,3}$ (90%, [α], +1.5°). Reduction (3 atm H₂/Pd-black/MeOH, 20°) of 13 followed by successive catalytic oxidation <code> (O $_{\rm o}$ /Pt-black/NaHCO $_{\rm o}$ /H $_{\rm o}$ O, 80°)</code> and esterification $(CH_2N_2/Et_2O, 20^{\circ})$ gave the two epimers $\frac{1}{4}$ and $\frac{1}{4}$ (TLC, Rf 0.29 and 0.35, PhH-EtOAc 3 : 2),

which were separated by column chromatography to give the 8R-epimer μ_0^3 (52%, $[\alpha]_D$ +90°) and the 8S-epimer $\sqrt[12]{3}$ (27%, [a]_D +75°). These epimers were equilibrated with K₂CO₃ in MeOH (60°). This structural assignment is based on conversion of $\frac{1}{4}$ into the naturally derived $\frac{26}{5}$ (8Repimer) as presented below.

The Cll-Cl7 segment for the synthesis was prepared in a stereochemically unambiguous manner starting with the branched D-allofuranose $\frac{1}{6}$, $\frac{3}{9}$ which was easily available from Dglucose. Acetylation (Ac₂0/Py, 20°) of \downarrow 6 gave the crystalline acetate χ ³ (95%, mp 67°, $\lbrack \alpha \rbrack_{\text{n}}$ +65°), which was converted into lg^{3} [mp 117° (needles), $\lbrack \alpha \rbrack_{\text{p}}$ +53°] by a four-step sequence in 84% overall yield: (1) 90% AcOH, 50°; (2) NaIO,/90% MeOH, 20°; (3) Zn(BH,) $_2$ /Et $_2$ O, 0°; (4) TsCl/Py, 20°. Treatment of λ 8 with MeMgBr $^{\circ}$ (L1,CuCl,/THF, 50°, 4 h) afforded λ 9 (99%, $\lceil \alpha \rceil_p$ +72°). Hydrolysis of \lg (2.5% H₂SO₄/t-BuOH, 50°) followed by oxidation by Br₂ (70%) dioxane, 20°, 48 h) to give the γ -lactone, which was treated with trityl chloride (Py, 20°) to give $2\alpha^3$ (70%, mp 142°, [a]_D -10°). Reaction of 2α with MeLi (Et₂0, -78°, 2 h) to afford $2\alpha^3$ (92%, $[\alpha]_n +8^\circ$), which was submitted to Wittig reaction (Ph₃P=CHCOOMe/PhMe, 80°, 72 h) to give χ^3 (76%, $\begin{bmatrix} \alpha \end{bmatrix}_{D}$ +20°). Silylation (t-BuMe₂SiCl/imidazole/DMF, 60°, 20 h) followed by reduction $(LiAlH_4/Et_20, 20^\circ)$ afforded 23^3 (93%, $[\alpha]_D^2$ +50°). Hydroboration¹¹ (BH₃-Me₂S/CH₂C1₂, 20°, 72h; aq. EtOH/NaOH/H₂O₂, 20°, 2 h) of 2λ gave the 1,2-diol which upon glycol cleavage (NaIO₄/90%) MeOH, 0°) afforded an aldehyde. This was treated with MeONa (MeOH, 20°) to give the E unsaturated aldehyde $\chi^{3,12}$ (57% from χ), [α]_D +4°) with ß-elimination, which was desilylated by Bu,NF (THF, 20°) to give the aldehyde $25^3.$

Thus, the epimer 14 was subjected to the aldol condensation with 25 (LDA/THF, –78° to 0°, 2 h) as previously described to give the desired E , E -unsaturated ketone 26 $(41\% , [\alpha]_{n}$ +66 $^{\circ}$, $\lambda_{\perp}^{\rm new}$ 286 nm (e 22200)). The other epimer 15 was not converted into 26 by the same procedure but into the 8S-epimer $\chi\chi^2$ ([$\alpha]_{_{\rm D}}$ +44°, Rf 0.54 and 0.49 for $\chi\beta$ and $\chi\chi$ on TLC with hexane-EtOAc 2 : 3). Synthetic $26/26$ was identical in all respects with a sample of the same structure produced by an unambiguous series of transformations (outlined below) starting from a naturally derived aglycone $2a$. Reduction (NaBH₄/MeOH, 0°) of 26 gave the C9-OH epimers $28a$ (48%, $[\alpha]_D$ +28°, Rf 0.38 on TLC with hexane-EtOAc 3 : 2) and $28R$ (42%, $[\alpha]_D$ +40°, Rf 0.34), which were saponified (0.2N KOH/aq. MeOH, 60°, 15 h) to afford the corresponding seco acids (~85%). The two acids were converted with 2,2'-dipyridyl disulfide (Ph₃P, 20°)¹³ into the 2-pyridinethiol esters $29a^3$ (Rf 0.63 on TLC, PhH-EtOAc $1\,$: 2) and $29b^3$ (Rf 0.58) respectively, which were submitted to cyclization $\tilde{}$ (PhMe, 110° , 20 h) to afford the 16 –membered–ring lactones $\rm 30a^{-1}$ (41%, mp ~94° (amorphous), $\left[\alpha\right]_n$ +37°, $\lambda_{\rm max}^{\rm max}$ 232 nm (e 33900), Rf 0.35 on TLC with hexane-EtOAc 3 : 2) and $30b^3$ (28%, mp 181° (needles), $[\alpha]_p$ +65°, $\lambda_{\text{max}}^{\text{mean}}$ 231 nm (E 32700), Rf 0.50), respectively. Selective oxidation (CrO $_2/$ HMPT, 20 $^{\circ}$, 20 h) of 30a and 30b provided the desired aglycone 2a (86%, mp 124° (needles), $[\alpha]_n$ +60°, $\lambda_{\rm max}^{\rm max}$ 282 nm (ε 21600)) identical in all respects with a naturally derived authentic sample. By this completion, the absolute configuration (especially at C14)^{2b} of tylosin $(\frac{1}{\lambda})$ was confirmed.

Treatment of a tylonolide hemiacetal, which was prepared from $\frac{1}{k}$, with Amberlyst 15 (H type) in MeOH (20°) gave the corresponding methyl furanosides, 2c which were tritylated (TrCl/Py, 50°) to afford two anomers λ_0^3 (65%, mp 124°, [α]_D +60°, Rf 0.56 on TLC with hexane-EtOAc 3 : 2) and λ_0^{3} (22%, mp ~85°, $[\alpha]_D$ 0°, $[\alpha]_{365}$ -83°, Rf 0.60). Reduction (NaBH₄/MeOH, 20°) of λ_0 to give $\frac{20}{30}$, followed by successive saponification (KOH/aq. MeOH, 60°, 15 h) and esterification (CH₂N₂/Et₂O, 20°) gave 28a (64%) and 28b (27%), which were selectively oxidized (MnO₂/CH₂C1₂, 20°, 48 h) to afford a reference sample of 26 (79%). Treatment of $2a$ with K_2CO_3 in MeOH (70°, 20 h) resulted in epimerization at C8 to give directly a mixture of $\lambda \in (41\%)$ and $\lambda \in (40\%)$.

With the synthesis of λ_{α} completed, the conversion into tylosin (1) can now be addressed. Acknowledgment: We are grateful to the Institute of Microbial Chemistry for the generous support of our program. We also are indebted to Mr. H. Takahashi for his technical assistance and Ms. R. Kobayashi of the Institute for NMR (25OMHz) spectral analyses.

References and Notes:

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tions established by mass spectra and/or elemental analyses. Melting points were uncortions established by mass spectra and/or elemental analyses. Melting points were uncorrected. Optical rotations were done in CHCl, at c 1.00 (20°) unless stated otherwise. NMR (δ , ppm from TMS, and J in Hz) spectra wĕre in CDCl, solution. Significant NMR spectral data are the following. 2a: 3.34(s, MeO), 5.02(m, H-6" & 15), 5.95(d, J=IO.5, H-13),
6.35(d, J=16, H-10). 2b: 3.35(s, MeO), 4.93(d, J=6, H-6"), 5.02(dt, J=10 & 3, H-15). 4: 0.95(d, J=7.5, Me), 4.10(dd, J=9 & 4, H-4). 5: 4.60(dt, J=7 & 4, H-4), 9.73(sharp t, CHO). 7: 1.40(s, acetonide), 6.06(dd, J=13 & 2, H−6'), 6.81(dd, J=13 & 3, H−6). 8a: 0.84(d, J=7. Me-4), 2.38(s, MeS), 2.74(s, MeSO), 3.96(d, J=2, H-7). &b: 0.88(d, J=7, 2.74(s, MeSO), 3.96(d, J=3, H–7). 9a: 1.26(t, J=7.4, <u>Me</u>CH₂S), 2.15(s, MeS).
7.4, <u>Me</u>CH₂S), 2.21(s, MeS). 10a: 2.05(s, MeS), 3.20(s, MeO). 10b: 2.01(s, M 0). 11: 4,97(dd, J=6 & 3, H−6"), 9.53(d, J=3, CHO). 11': 4.91(t, J=3, H−6"), 9.46(d, J=3, CHO). 13: 2.23(s, Me-8), 6.51(dd, J=ll & 1, H-7). 14: 1.14(d, J=7, Me-8), 2.16(s, COMe), 3.72(s, COOMe). 15: 1.11(d, J=7, Me-8), 2.16(s, COMe), 3.72(s, COOMe). 18: 2.03(s, OAc),
2.42(s, Me of Ts). 20: 0.95(t, J=7, Me), 1.62(quintet, C<u>H</u>₂Me). 22: 2.03(s, vinyl Me), 3.60 (s, COOMe), 5.88(s, vinyl H). $\mathcal{Z}_{\mathcal{A}}^{A}$: -0.10 & 0.00(each s, Me $_{2}$ Si), \mathbf{S} (s, COOMe), 5.88(s, vinyl H). 24: -0.10 & 0.00(each s, Me₂Si), 1.13(s, t-BuSi), 1./5(s, Me-
12), 6.40(d, J=11, H-13), 9.34(s, CHO). 25: 6.57(d, J=11, H-13), 9.33(s, CHO). 26: 0.94(d, J=6.5, Me-4), l.l5(d, J=7, Me-8), 6.l3(d, J=10, H-13), 6.l4(d, J=16, H-10). $\chi\!\!\!\chi$: 0.92(d, J= 6.5, Me-4), 1.14(d, J=7, Me-8). 28a: 4.00(m, H-9), 5.59(dd, J=15.5 & 7, H-10), 5.65(d, J=
10, H-13). 28b: 4.16(m, H-9), 5.61(dd, J=15.5 & 7, H-10), 5.63(d, J=8.5, H-13). 29a: 5.55 (dd, J=16 & 8, H−10), 5.63(d, J=10, H−13), 8.51(sharp m, SPy-∐). 29b: 5.62(dd, J=16 & 7, H− 10), 5.62(d, J=9.5, H-13), 8.59(sharp m, SPy-<u>H</u>). 30a: 4.27(dull dt, H-9), 4.99(m, H-6" & 10), 5.62(d, J=9.5, H-13), 8.59(sharp m, SPy-H). 30a: 4.27(dull dt, H-9), 4.99(m, H-6" & 15), 5.77(dd, J=16 & 9, H-10). 30a: 4.25(sharp m, H-9), 4.99(m, H-6" & 15), 5.77(dd, J=16 & 3.5, H-10).
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J=4, H-1); the $\lbrack \circ \rbrack \; \lbrack \alpha \rbrack \; \rbrack \; \gamma \; \rbrack \; \gamma \; \rbrack \; \; \alpha \; \lbrack \alpha \rbrack \; \rbrack \; \; \alpha \; \lbrack \alpha \rbrack \; \rbrack \; \; \alpha \; \lbrack \alpha \; \lbrack \, \rbrack \; \; \gamma \; \rbrack \; \gamma \; \gamma \; \rbrack \; \gamma \; \gamma \; \r$ H-45, 4.19(dd, J=10.5 & 9.5, H-3'a), mp 109-110°, $[\alpha]_{\overline{D}}$ +67°, NMR: 2.3(m, H-3), 3.6(m, 4.50(dd, J=9.5 & 4, H-3'b). 4.63(t, J=4, H-2), 5.74 (d, 524, H-l). The NMR data were identical with those reported in a literature: A. Rosenthal and M. Sprinzl, Can. *J. Chem., 47, 4477 (1969).*
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